

COBB2035 • RECITATION 2 (but actually 1)

PyMOL

Today

Install

Get open-source PyMOL running in a clean conda environment.

Demo

Follow Along

Basic ✓

Ray-traced figure of profilin 1: wild type vs. the M114T ALS variant.

Extra ✓+

Cryo-EM map of a glycine receptor. Identify evidence of a chloride atom that is not in the pdb.

Installing PyMOL

Open-source PyMOL. No license, no registration.

```
conda create -n pymol python=3.11
conda activate pymol
conda install conda-forge::pymol-open-source
```

Launch it:

```
pymol
```

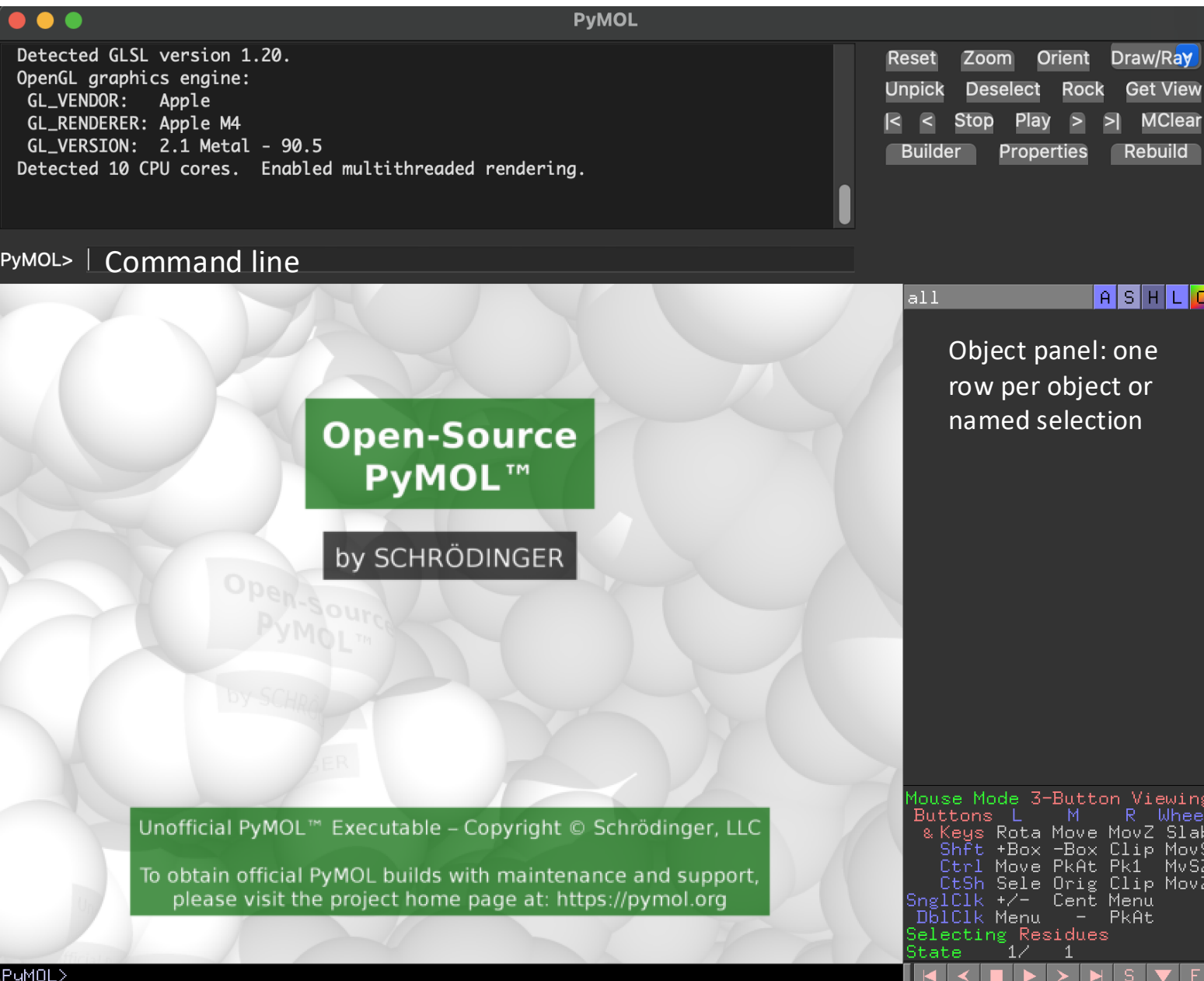
WHY A SEPARATE ENVIRONMENT

Cleaner to have separate tools in separate environments

IF THE SOLVER HANGS

- Use mamba, or conda install --solver=libmamba.

The window



Menus — everything here has a command equivalent

Object panel toggles:
Action, Show, Hide, Label, Color

Mouse Control Guide

Selection tools

Mouse > Selection Mode switches clicking between Atoms / Residues / Chains / Molecules.

Anatomy of a command

```
color    red,    resi 44 and chain A
#      ^      ^      ^
#  cmd    arg1   arg2
```

- Arguments are separated by commas, not spaces.
- Most commands take an optional selection as the last argument. If not present the command applies to everything.
- Keyword arguments come last: `png out.png, width=2000, ray=1`

The wiki helps: pymolwiki.org

Selections

Half of PyMOL is knowing how to say “these atoms and no others.”

PROPERTIES

<code>resi 114</code>	residue number
<code>resn MET</code>	residue name
<code>name CA</code>	atom name
<code>chain A</code>	chain
<code>elem S</code>	element
<code>ss H</code>	secondary structure
<code>b > 60</code>	B-factor

CLASSES

<code>polymer</code>	protein + nucleic acid
<code>solvent</code>	water
<code>organic</code>	small-molecule ligands
<code>inorganic</code>	ions, metals
<code>backbone</code>	and sidechain — within polymer
<code>hydro</code>	hydrogens

OPERATORS

<code>and or not</code>	Logic. and narrows, or widens.
<code>byres</code>	Grow to whole residues. Use it or you get half a sidechain.
<code>s1 within 5 of s2</code>	Atoms of s1 that are near s2
<code>s1 expand 5</code>	s1 plus its neighbours
<code>s1 around 5</code>	The neighbours only — s1 itself excluded

1YCR

```
fetch 1YCR, async=0
```

```
hide everything  
show cartoon, polymer  
color grey70
```

```
# colour N-to-C so you can read directionality  
spectrum count, blue_white_red, polymer and name CA
```

```
# Select residues, name the selected residues patch  
select patch, resi 8+44+68+70  
show sticks, patch and sidechain  
color orange, patch
```

```
orient
```



Representations and colour

SHOW / HIDE

show, hide, and as.
as replaces everything else

```
as cartoon  
show surface, polymer
```

```
# surfaces only:  
set transparency, 0.4  
set surface_color, grey70
```


Two structures: adenylate kinase

4AKE E. coli adenylate kinase, no ligand, open.

1AKE The same enzyme clamped shut around AP5A, a two-substrate mimic.

```
fetch 4ake, opened, async=0
fetch 1ake, closed, async=0
```

4AKE has two copies in the asymmetric unit. 1AKE has one. Split first:

```
# Extract chain A from opened
# Extract chain B from closed
# Delete initial structures
```

```
as cartoon
color grey70, chA_opened
color skyblue, chB_closed
```

```
align chA_opened, chB_closed
orient
```



`align`

Sequence alignment, then fit, then outlier rejection.
Default for near-identical sequences.

`super`

Structure-driven. Use when sequence identity is low.

Ray tracing and saving



```
bg_color white
set ray_opaque_background, 0
set antialias, 2
set ambient_occlusion_mode, 1
set light_count, 8
set ray_trace_mode, 0

png figure1.png, width=2000, height=1500, dpi=150, ray=1
```

`width, height`

`dpi=300`

`ray=1`

`bg_color`

`ray_opaque_background`

`ambient_occlusion_mode`

`ray_trace_mode`

Pixels. These are what determine resolution.

Metadata, so layout software scales it right.

Ray-traces first

The viewport you work in.

The file you export. 0 gives a transparent PNG.

Darkens crevices. Big readability win on surfaces.

0 photoreal, 1 adds black outlines, 2 line art, 3 cel-shaded.

Reference

Every wiki page has worked examples. Read them instead of guessing.

[Main Page](#)

Start here

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Pull from the PDB

[Load](#)

Local files and maps

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Representations

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Named colours, spectrum

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Sequence-based fit

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Structure-based fit

[Distance](#)

Measure, find H-bonds

[Ray](#)

Rendering

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.pml and Python

Cheat sheet

LOAD

<code>fetch lubq, async=0</code>	from the PDB
<code>load file.pdb, name</code>	from disk
<code>load map.map, edmap</code>	density
<code>delete name</code>	remove

LOOK

<code>hide everything</code>	reset
<code>as cartoon, polymer</code>	replace reps
<code>show sticks, sel</code>	add rep
<code>color grey70, sel</code>	
<code>orient sel</code>	rotate + zoom
<code>zoom sel, 5</code>	zoom only

COMPARE

<code>align a, b</code>	by sequence
<code>super a, b</code>	by structure
<code>align a, b, transform=0</code>	measure only
<code>distance d, s1, s2, 3.5, mode=2</code>	H-bonds

RENDER

`bg_color white`

`png f.png, width=2000, ray=1` always ray=1

DEBUG

`help <cmd>`

`print(cmd.count_atoms("sel"))`

`iterate sel, print(resi, resn)`

`get_view` camera

Profilin 1: Wild Type vs. M114T

4X1L Wild-type profilin 1

4X25 M114T profilin 1 — an ALS-linked variant

Make a **ray-traced** figure showing any differences between the two proteins. Write a caption. Put your name on it.

SUBMIT

Upload your figure(s) with a caption and your name to GradeScope.

A cryo-EM map, and a missing ion

- 1 Visualise the density map for 7M6Q.
Coordinates from PDB; Map from EMDB.
- 2 Find evidence of a chloride ion.
- 3 Comment on anything else you notice.

KNOW WHAT YOU ARE LOOKING AT

7M6Q is a full-length $\alpha 1$ **glycine receptor**, pentameric, in a lipid nanodisc, with glycine and THC bound. The map is **EMD-23704**.

The glycine receptor is an anion channel. Look into the channel.